

## THE BIOSYNTHETIC ORIGIN OF FUSICOCCIN HYDROXYISOPROPYL GROUP†

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**Abstract**—The biosynthetic origin of the hydroxyisopropyl group of fusicoccin **1** has been determined. Two out of the eight tritium mevalonoid atoms retained in **1** following incorporation of MVA-[2-<sup>3</sup>H<sub>2</sub>, 2-<sup>14</sup>C] have been located at C-20; whilst none has been found in the C-19 methylene group.

Studies on fusicoccin<sup>1</sup> **1** biosynthesis, performed by a British<sup>2</sup> and an Italian<sup>3</sup> group, have (a) demonstrated that fusicoccin is a diterpenoid, not a rearranged sesterterpenoid, (b) elucidated the mechanism of geranylgeranyl pyrophosphate cyclization implying consecutive 1,2-hydride shifts, (c) discriminated minor fusicoccins that are true intermediates in the biosynthetic pathway from those which represent shunt products.

The biosynthetic origin of HOH<sub>2</sub>C-19 and H<sub>3</sub>C-20, corresponding to the terminal methyl groups of the prenyl pyrophosphate, has remained an open question; in this paper we show that the methyl group is entirely derived from C-2 of mevalonic acid.

### RESULTS AND DISCUSSION

A previous study<sup>3a</sup> had demonstrated that six out of the eight tritium atoms retained in **1**, following incorporation of MVA-[2-<sup>3</sup>H<sub>2</sub>, 2-<sup>14</sup>C], are located in the aglycone moiety. In particular, one tritium was found at C-8<sup>3a</sup> and one at C-12;<sup>3b</sup> according to the isoprenoid rule, two are expected to be present at C-4 and two at either C-19 or C-20. Labelled **1**, prepared by feeding cultures of *Fusicoccum amygdali* Del. with MVA-[2-<sup>3</sup>H<sub>2</sub>, 2-<sup>14</sup>C], was diluted with inactive material and chemically degraded to give the derivatives reported in Scheme 1. In two independent experiments, **1** was transformed, through known reactions,<sup>1</sup> into the aglycone and the corresponding 8,9-isopropylidene derivative **2**.

In a first experiment, **2** was oxidized, by pyridinium chlorochromate, to the rather unstable 12,19-dioxoderivative **3** which on reduction by NaBH<sub>4</sub> afforded a mixture of **2** and its 12-epimer **4**; this was fractionated by column chromatography and the epimers separately counted as 12,19-dimesyl derivatives **6**<sup>3b</sup> and **5**, respectively (Scheme 1, route a).

The <sup>3</sup>H:<sup>14</sup>C ratios of **5** and **6** (Table 1) are consistent with the loss of one tritium atom on oxidation of **2**; the loss is from C-12, since it was previously demonstrated<sup>3b</sup> that the 19-trityl derivative of **2**,<sup>1c</sup> prepared from fusi-

coccin biosynthesized from MVA-[2-<sup>3</sup>H<sub>2</sub>, 2-<sup>14</sup>C], loses one <sup>3</sup>H when it reacts with pyridinium chlorochromate to afford the corresponding 12-oxo-compound.<sup>3b</sup> In conclusion the presence of <sup>3</sup>H at C-19 is ruled out.

In a second experiment, **1** was converted, through known reactions,<sup>3b</sup> into the tetraene **7**. Oxidation of **7** with SeO<sub>2</sub> gave compound **8** (14% yield) which afford a monoacetate **10**. Spectroscopic data of compounds **8** and **10** allow the assignment of a 20-hydroxytetraene structure to **8**. Treatment of **8** with activated MnO<sub>2</sub> resulted in its quantitative conversion to the α,β-unsaturated aldehyde **9** (Scheme 1, route b). As shown in Table 1, 1.05 tritium atoms are lost on passing from **6** to **9**. This result is in reasonable agreement with the theoretical value of 1½. In fact, hydroxylation of the C-20 methyl results in the statistical removal of ½ of a tritium atom and the subsequent oxidation of HOH<sub>2</sub>C-20 to the aldehyde causes a further 50% loss of 1½ atoms.

The results of the degradations clearly show that two of the hydrogen atoms at C-20, and none at C-19, are derived from C-2 of mevalonate. Thus, the terminal methyl groups of the geranylgeranyl pyrophosphate retain their individuality in the formation of **1**, as demonstrated for other polycyclic diterpenes,<sup>4</sup> sesquiterpenes<sup>5</sup> and triterpenes.<sup>6</sup>

### EXPERIMENTAL

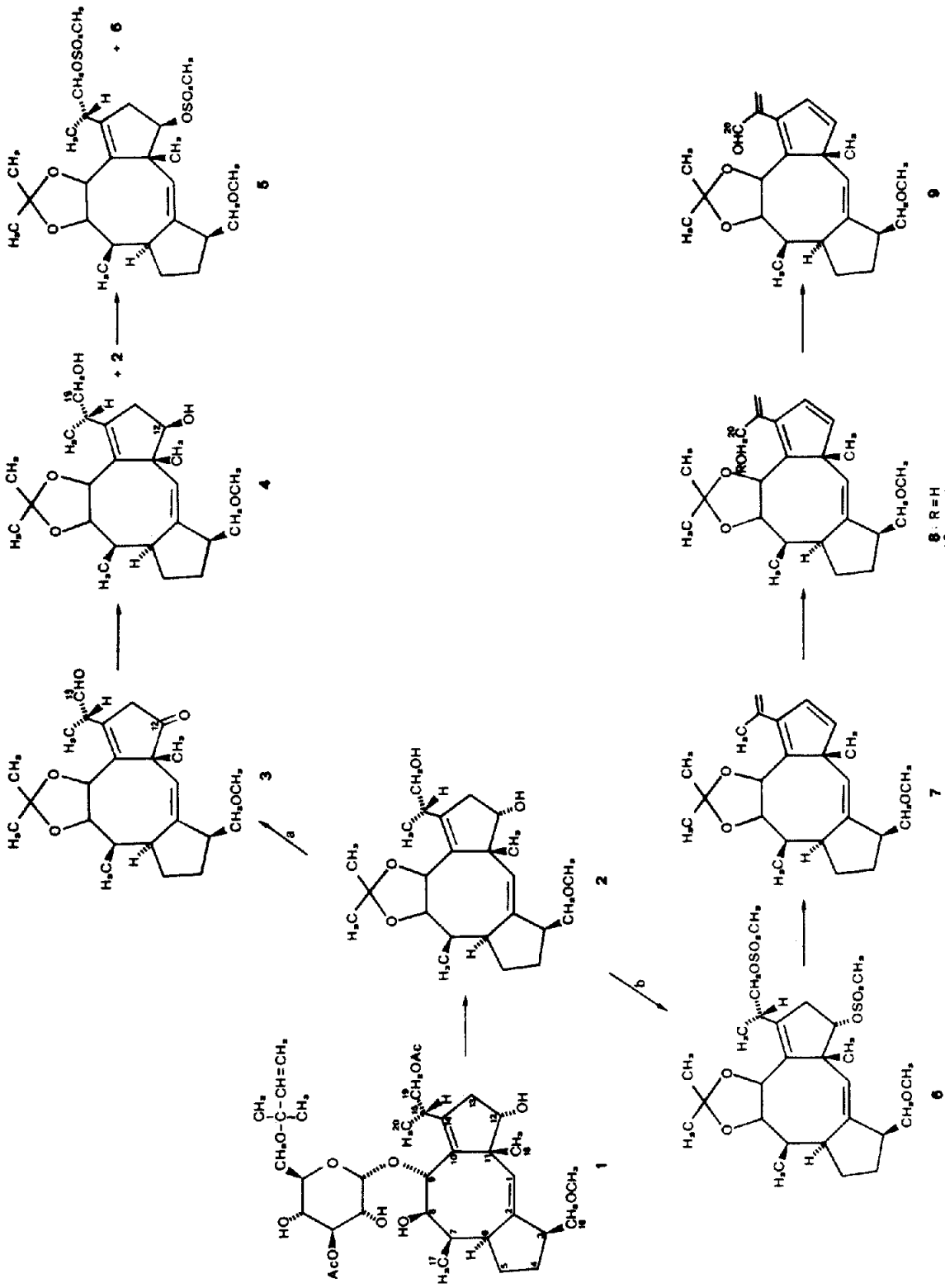
*General methods.* Melting points are uncorrected. Optical rotations were measured on a Perkin-Elmer 141 polarimeter in CHCl<sub>3</sub> solutions. IR spectra were recorded on a Perkin-Elmer

Table 1. Radioactivity (<sup>3</sup>H) retained in some derivatives prepared from fusicoccin isolated from cultures of *Fusicoccum amygdali* Del. supplemented with MVA-[2-<sup>3</sup>H<sub>2</sub>, 2-<sup>14</sup>C]

Compound	Experiment	<sup>3</sup> H/ <sup>14</sup> C	Atom ratio
<b>2</b>	1	6.45	6.00*
<b>5</b>	1	5.33	4.96
<b>6</b>	1	5.14	4.78
<b>6</b>	2	4.45	6.00*
<b>9</b>	2	3.67	4.95

\*Theoretical values based on previous results.<sup>3a</sup>

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6: R=H  
10: R=Ac  
Scheme 1. Degradation of fuscococcin-[<sup>3</sup>H, <sup>14</sup>C].

399 instrument for solutions in  $\text{CHCl}_3$ . UV spectra were measured for solutions in  $\text{CH}_3\text{CN}$  or EtOH on a Varian Cary 210 spectrophotometer.  $^1\text{H}$  NMR spectra were recorded at 270 MHz on a Bruker spectrometer in  $\text{CDCl}_3$  using TMS as internal standard; chemical shifts are in ppm. Mass spectra were recorded on an AEI-902 mass spectrometer at 70 eV. Analytical and preparative TLC were performed on silica gel plates (Merck, Kieselgel 60 F<sub>254</sub> 0.25 and 2 mm, respectively) and spots were developed by spraying with a 10%  $\text{H}_2\text{SO}_4$  solution in MeOH and then heating at 105°C. Column chromatography was carried out on silica gel (Merck, Kieselgel 60 0.063–0.2 mm).

**Feeding and product isolation.** Fermentations were carried out according to a published procedure<sup>7</sup> scaled down to 500 ml Erlenmeyer flasks containing 100 ml of medium. Specific activities of the labelled precursors were the highest commercially available (Amersham). Labelled **1**, prepared from two independent fermentations and separately used for experiments 1 and 2, was extracted with  $\text{CHCl}_3$ , diluted with unlabelled **1** and purified by column chromatography to give about 1220 and 187 dpm  $\text{mg}^{-1}$  (batch 1), 952 and 215 dpm  $\text{mg}^{-1}$  (batch 2) for  $^3\text{H}$  and  $^{14}\text{C}$ , respectively. Radioactivity measurements were carried out with a Beckman LS 8100 liquid scintillation counter, using a solution containing 6.5 g of PPO, 130 mg of POPOP and 104 g of naphthalene in 1000 ml of toluene-dioxane (1:1 v/v) as scintillant. All compounds were crystallized to constant specific activity.

**12,19-dioxo-8,9-isopropylideneaglycone of fusicoccin 3.** Pyridinium chlorochromate<sup>8</sup> (4.73 g) was added to a soln of **2** (150 mg) in dry  $\text{CH}_2\text{Cl}_2$  (10 ml); the mixture was stirred at room temp. and monitored by TLC ( $\text{CHCl}_3$ -isopropanol 95:5 v/v). After 4 h, when all the starting material had reacted, the reaction was stopped with anhydrous Et<sub>2</sub>O (20 ml). The mixture was filtered through a short  $\text{SiO}_2$  column which was then washed with the same solvent. The combined filtrates and washings were evaporated under reduced pressure. Purification of the residue by preparative TLC ( $\text{CHCl}_3$ -isopropanol 95:5 v/v) afforded an oil (103 mg, 68% yield) which crystallized from pet. ether (b.p. 40–70°): m.p. 115–118°;  $[\alpha]_D^{25} = -42.3$  ( $c = 0.95$ ); UV ( $\text{CH}_2\text{CN}$ ),  $\lambda_{\text{max}}(\epsilon)$ : 294 nm (640); IR,  $\nu_{\text{max}}$ : 2740, 1750, 1730  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR  $\delta$ : 0.81 (3H, d,  $J = 7$  Hz, 20-Me), 1.1 (3H, d,  $J = 7$  Hz, 17-Me), 1.20 (3H, s, 18-Me), 1.27 and 1.39 (3H each, s, two Me of isopropylidene group), 2.45 (1H, d,  $J = 24$  Hz, 13-H), 2.87 (1H, d,  $J = 24$  Hz, 13-H), 3.20 (3H, s, OMe), 3.80 (1H, dd,  $J = 10$  and 5 Hz, 8-H), 4.15 (1H, m, 15-H; 1H, m, 9-H; by  $^1\text{H}$  NMR), 5.25 (1H, t,  $J = 1.5$  Hz, 1-H) and 9.4 (1H, s, 19-H); MS  $m/e$ : 402 (25%,  $M^+$ ), 373 (50), 344 (50), 315 (100) and 283 (81).

**12-epi-8,9-isopropylideneaglycone of fusicoccin 4.** Compound **3** (103 mg) in MeOH (10 ml) was stirred with  $\text{NaBH}_4$  (206 mg) at room temp. TLC ( $\text{CHCl}_3$ -isopropanol 95:5 v/v) showed that after 30 min all the starting material had been reduced. After decomposition with 0.1N HCl of borohydride excess, the mixture was diluted with water (200 ml) and extracted with  $\text{CH}_2\text{Cl}_2$  (3 × 60 ml). The combined extracts were washed with water and dried ( $\text{Na}_2\text{SO}_4$ ). Evaporation of the solvent gave an oily residue consisting of **2** and its 12-epiderivative **4**. Column chromatography of the mixture eluted with  $\text{CHCl}_3$ -isopropanol 95:5 v/v afforded **2** (29 mg, 33% yield) followed by **4** (48 mg, 47% yield); the latter crystallized from Et<sub>2</sub>O-*n*-hexane: m.p. 199–200°;  $[\alpha]_D^{25} = -33.9$  ( $c = 4.41$ ); UV (EtOH),  $\lambda_{\text{max}}$ : < 220 nm; IR,  $\nu_{\text{max}}$ : 3450, 1600  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR,  $\delta$ : 0.92 (3H, d,  $J = 7$  Hz, 17-Me), 0.96 (3H, d,  $J = 7$  Hz, 20-Me), 1.12 (3H, s, 18-Me), 1.35 and 1.54 (3H each, s, two Me of isopropylidene group), 2.11 (1H, dd,  $J = 18$  and 6 Hz, 13-H), 2.57 (1H, dd,  $J = 18$  and 6 Hz, 13-H), 3.35 (3H, s, OMe), 3.55 (1H, m, 15-H), 3.95 (1H, m, 8-H; 1H, m, 12-H; by  $^1\text{H}$  NMR), 4.18 (1H, d,  $J = 9$  Hz, 9-H), and 5.54 (1H, t,  $J = 1.5$  Hz, 1-H); MS  $m/e$ : 406 (5.4%,  $M^+$ ), 391 (1.5), 388 (3.6), 348 (13), 330 (31) and 310 (100).

**12-epi-12,19-dimesyl derivative of 8,9-isopropylideneaglycone of fusicoccin 5.** The compound **5** was prepared from **48** mg of **4** according to a reported procedure.<sup>3b</sup> The reaction afforded 67 mg of a solid (99% yield) which crystallized from  $\text{Me}_2\text{CO}$ -pet. ether (b.p. 40–70°): m.p. 158–159°;  $[\alpha]_D^{25} = -14.6$  ( $c = 5.13$ ); UV ( $\text{CH}_3\text{CN}$ ),  $\lambda_{\text{max}}$ : < 220 nm; IR,  $\nu_{\text{max}}$ : 3450, 1600  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR,  $\delta$ : 0.94 (3H, d,  $J = 7$  Hz, 17-Me), 1.12 (3H, d,  $J = 7$  Hz, 20-Me), 2.46 (1H, dd,  $J = 15$  and 6 Hz, 13-H), 2.82 (1H, dd,  $J = 15$  and 6 Hz,

13-H), 3.04 (3H, s, O<sub>2</sub>SMe), 3.06 (3H, s, O<sub>2</sub>SMe), 3.39 (3H, s, OMe), 3.92 (1H, dd,  $J = 10$  and 6 Hz, 8-H), 4.77 (1H, dd,  $J = 6$  Hz, 12-H). The other proton resonances are very similar to those of **6**; MS,  $m/e$ : 562 (8.8%,  $M^+$ ), 547 (1.6), 530 (0.8), 504 (3.5), 472 (8.8), 466 (26), 451 (13) and 79 (100).

**20-hydroxytetraene 8.** A soln of **7** (310 mg) in EtOH (51 ml) was heated under reflux with  $\text{SeO}_2$  (993 mg); the reaction was followed by TLC (*n*-hexane - EtOAc 7:3 v/v). After 3.5 h the mixture was poured into H<sub>2</sub>O (250 ml) and then extracted with Et<sub>2</sub>O (3 × 100 ml). The combined extracts, after washing with a saturated  $\text{NaHCO}_3$  solution followed by water, were dried ( $\text{Na}_2\text{SO}_4$ ). Evaporation of the solvent gave a brown oil containing at least four chromatography distinguishable compounds. Column chromatography (*n*-hexane-EtOAc 7:3 v/v) of the mixture separated the main component from unreacted material and minor reaction products. Further purification by preparative TLC, eluted with same solvent, afforded a crystalline solid (44 mg, 14% yield): m.p. 86–88°;  $[\alpha]_D^{25} = +120.6$  ( $c = 1.24$ ); UV (EtOH),  $\lambda_{\text{max}}(\epsilon)$ : 274 nm (1987); IR,  $\nu_{\text{max}}$ : 3420, 1610  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR,  $\delta$ : 0.95 (3H, d,  $J = 7$  Hz, 17-Me), 1.36 (3H, s, 18-Me), 1.39 and 1.47 (3H each, s, two Me of the isopropylidene group), 2.06 (1H, d,  $J = 7$  and 4.4 Hz 7-H), 2.64 (1H, m, 3-H; 1H, m, 6-H; by  $^1\text{H}$  NMR), 3.36 (3H, s, OMe), 4.13 (1H, dd,  $J = 9.6$  and 4.4 Hz, 8-H), 4.22 (2H, m, 20-H), 4.48 (1H, d,  $J = 9.6$  Hz, 9-H), 5.06 (1H, d,  $J = 1.8$  Hz, 19-H), 5.27 (1H, d,  $J = 1.8$  Hz, 19-H), 5.52 (1H, t,  $J = 1.8$  Hz, 1-H), 6.06 and 6.27 (1H-12 and 1H-13, d,  $J = 5.5$  Hz); MS,  $m/e$ : 386 (94%,  $M^+$ ), 368 (26), 343 (28), 328 (47) and 311 (100).

**20-oxotetraene 9.** The compound **8** (44 mg) in dry *n*-hexane (35 ml) containing active  $\text{MnO}_2$  (250 mg) was stirred at room temp. The reaction was monitored by TLC (*n*-hexane-EtOAc 7:3 v/v); after 4 h all **8** was reacted. The mixture was filtered through Celite and this was washed with *n*-hexane (300 ml). The oily residue left after evaporation of the combined filtrates was purified by preparative TLC (*n*-hexane-EtOAc 7:3 v/v) to afford a crystalline solid (41 mg, 94% yield) which was recrystallized from *n*-hexane: m.p. 104–105°;  $[\alpha]_D^{25} = +42.2$  ( $c = 0.45$ ); UV, (EtOH),  $\lambda_{\text{max}}(\epsilon)$ : 274 nm (2057); IR,  $\nu_{\text{max}}$ : 1705  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR,  $\delta$ : 1.27 and 1.37 (3H each, s, two Me of the isopropylidene group), 1.35 (3H, s, 18-Me), 2.62 (1H, m, 3-H; 1H, m, 6-H; by  $^1\text{H}$  NMR), 3.84 (1H, dd,  $J = 9.9$  and 4.4 Hz, 8-H), 4.48 (1H, d, 9.9 Hz, 9-H), 6.25 (1H, d,  $J = 1.8$  Hz, 19-H), 6.45 (1H, d,  $J = 1.8$  Hz, 19-H), 9.62 (1H, s, 20-H); the other proton resonances are very similar to those of **8**; MS,  $m/e$ : 384 (59%,  $M^+$ ), 339 (41), 327 (100) and 281 (35).

**20-acetyltetraene 10.** Acetylation of **8** (4.6 mg) with  $(\text{CH}_3\text{CO})_2\text{O}/\text{Py}$  afforded the 20-acetyl derivative; the usual work-up gave an oil (5.2 mg); IR,  $\nu_{\text{max}}$ : 1725  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR,  $\delta$ : 2.05 (3H, s, OMe), 4.76 (2H, m, 20-H), 5.30 (1H, d,  $J = 1.8$  Hz, 19-H), 5.34 (1H, d,  $J = 1.8$  Hz, 19-H); the other proton resonances are very similar to those of **8**.

**Nomenclature** - 12,19-dioxo-8,9-isopropylideneaglycone of fusicoccin **3**. Dicyclopenta [3,4:6,7] cycloocta [1,2-d] - 1,3-dioxole - 4 - ethanal - 6(3aH) - one - 5,6a,8,9,10,10a,11,11a-octahydro - 8 - (methoxymethyl) -  $\beta$ ,2,2,6a,11-pentamethyl. [3aR - [3aa, 4(S\*),6a $\beta$ ,8 $\beta$ ,10aa,11 $\beta$ ,11a $\beta$ ]]. **20-oxotetraene 9**: Dicyclopenta [3,4:6,7] cycloocta [1,2-d] - 1,3-dioxole - 3a,6a,8,9,10,10a,11,11a - octahydro - 8 - (methoxymethyl) - 2,2,6a,11-tetramethyl - 4 - (1-al - ethenyl). [3aR[3aa,(S\*),6a $\beta$ ,8 $\beta$ ,10aa,11 $\beta$ ,11a $\beta$ ]].

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